Adipose Tissue-Resident Macrophages and Obesity

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Author's Contribution

All the authors contributed significantly to the research that resulted in the submitted manuscript.

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ABSTRACT

Type 2 diabetes and insulin resistance are one of the major consequences of obesity as a result of inflammation in adipose tissues. Adipose tissue-resident macrophages (ATMs) have a primary role in tissue remodeling and maintenance of homeostasis within adipose tissue. Two different types of macrophages are present in adipose tissue i.e. M1 and M2-type. Obesity is associated phenotypic transformation of macrophages, from anti-inflammatory M2 to pro-inflammatory M1 macrophages. M1-type macrophages increases in obesity and contribute to the development of type 2 diabetes and insulin resistance. In contrast, M2 ATMs secretes anti-inflammatory cytokines and are involved in maintaining insulin sensitivity. However, little is known about the role of M2 macrophages in adipose tissue. Literature related to the role of M1 and M2 macrophages in metabolism have been reviewed in this article with emphasis on the macrophages associated with adipose tissues. Role of M2 macrophages in adipose tissues have also been highlighted in this article to enhance our knowledge and understanding of macrophages homogeneity.

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INTRODUCTION

Adipokines are the hormones that are responsible for systemic metabolism released by the adipose tissues. Health of adipokines is maintained via macrophages. Activated macrophages (M1) accumulate as the size of adipocytes becomes larger in adipose tissues and trigger systemic and local inflammation by worsening insulin resistance. Alternatively, activated macrophages (M2) possess anti-inflammatory actions. However, the exact mechanism how health of adipose tissues in a lean state is maintained remains elusive.

In obesity during pathological expansion, growth of neovasculature fails to catch up with rapid adipose tissue expansion resulting in adipose tissue hypoxia leading to adipose tissue dysfunction and systemic insulin

resistance [1, 2]. In contrast to healthy pathological expansion expansion, characterized by some prominent features macrophage including infiltration (inflammation), hypoxia and limited vessel growth [2]. Although hypoxia due to insufficient angiogenesis is likely to play a key casual role in the development of pathological expansion, mechanism underlying precise inadequate angiogenesis in pathological expansion and adequate angiogenesis in healthy expansion is not yet determined.

In obese adipose tissue, hypoxia has been shown to induce the stimulation of hypoxia-inducible factor-1 (HIF-1) [3, 4]. Hypoxic environment causes its HIF-1 α subunit to translocate from cytoplasm to the nucleus, where it dimerizes with its HIF-1 β subunit and certain co-activators interact with its

transactivation domain and results in the transcription of many genes regulating different processes including angiogenesis and glucose metabolism [5].

Hypoxic responses mediated by HIF-1α affect the adipocytes and macrophages differentially. In adipocytes HIF-1α has been shown to cause the metabolic dvsfunction followed bv proinflammatory response as reported in several transgenic mice studies [4, 6-10]. With regard to the role of macrophage HIF-1α we hypothesize that there are two possibilities about the dual character of macrophage HIF-1α. The first possibility is that macrophage HIF-1α is involved in proinflammatory responses [11, 12], thereby causing the adipose tissue inflammation and systemic insulin resistance. The other possibility is that adipose tissueresident macrophages (ATMs) secrete proangiogenic genes such as vascular endothelial growth factor (VEGF), basic fibroblast growth factor, as reported in tumorassociated macrophages (TAMs) [13] under hypoxic conditions, thus providing adipocytes with vascular network to improve adipocyte metabolism.

Adipose vasculature is regulated by various proangiogenic factors released from adipocytes and other cells in the stromal vascular fraction such as macrophages, preadipocytes and endothelial cells. Production of these factors may be impaired in pathological expansion of adipose tissue. There are several pieces of evidence pointing towards the positive role of adipocyte-derived VEGF in inducing healthy expansion [14]. Under the obese state, the expression of VEGF gene is not sufficiently induced [12], which may be partly due to the fact that adipocyte HIF-1α is not a strong inducer of VEGF gene expression. In addition, adipocyte HIF-1α not only failed in inducing the adequate angiogenesis but was involved in the induction of fibrosis through increased formation of extracellular matrix components (ECM) [4]. On the other hand, ATMs express

the VEGF gene via HIF-1α in response to adipose tissue hypoxia. Moreover, differentiating preadipocytes produce high level of a range of proangiogenic factors, including fibroblast growth factor 2 (FGF-2), VEGF, hepatocyte growth factor (HGF) and PDGFs [15]. An early study showed that differentiation from preadipocytes to mature adipocytes was concomitantly linked to elevated production of angiogenic factors [16]. Recently, we identified that macrophage HIF-1α prevents angiogenesis in preadipocytes [17] (Figure 1).

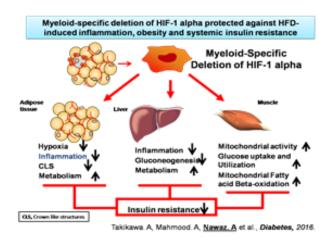


Figure 1. Deletion of HIF-1 α improve systemic insulin resistance.

Deletion of HIF-1α from myeloid cells causes significant low inflammatory response from adipose tissue-resident macrophages (ATMs), which then turn the expansion of adipose tissue from pathological to healthy one characterized by low fat cell mass, improved metabolism evident by upregulation of PPARy gene as well as adiponectin level in adipocytes along with reduction in fibrosis and oxidative stress, but keeping in view that these changes come as secondary changes to reduce proinflammatory responses in macrophages. This improved adipose tissue state was associated with improved insulin signaling in liver and skeletal muscle of KO mice. Of note, increased mitochondrial activity in skeletal muscle

and reduced expression of gluconeogenic genes in liver from HIF-1α KO mice was observed [17].

Collectively, our results demonstrate that HIF1 α inhibition in macrophages leads to significant metabolic improvements, suggesting that selective HIF1 α inhibition in macrophages may be an effective therapeutic target in the context of metabolic dysfunction.

Adipose tissue macrophage as in other organs pivotal role in the physiological homeostasis. Much has been known about macrophage role in inflammatory process in obese adipose tissue which is known closely related with the development of insulin resistance both in human and animal [18]. however, much less is known about their role in lean condition. Since majority of macrophage in lean adipose tissue is M2 type macrophages. we decided to focus our attention to their physiological role in that condition.

We previously demonstrated that majority of adipose tissue macrophages in lean mice are M2 types which could be identified and separated as distinct macrophage population using surface marker CD206 [19]. So, in subsequent study, we would like to extend our previous data to dissect the role of M2 macrophage using CD206 as marker. To obtain information about M2 macrophage role we used a conditional depletion method using diphtheria toxin-receptor ablation system. Diphtheria toxin receptor-mediated ablation is a specific cell ablation system in which diphtheria (DTR) toxin receptor is transgenically expressed specific cell targets. on Administration of diphtheria toxin (DT) enables the ablation and/or depletion of the targeted cells [20]. This technique has been used effectively in previous studies [20, 21]. This system relies on the fact that the mouse DTR binds DT poorly compared with the human molecule. Here, we elucidated the roles of M2 macrophages in adipose tissue using CD206

as a marker by employing diphtheria toxin receptor-mediated ablation system.

To our surprise, the result of this study shows us that M2 macrophage is pivotal in modulating adipocyte progenitors' proliferation and differentiation into adipocyte through TGF β signaling [22] (Figure 2). We anticipate that CD206 may provide a niche for adipocyte progenitors to keep them in hibernation state.

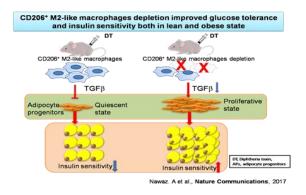


Figure 2. Depletion of CD206 M2 macrophages promote adipocyte progenitor's proliferation and induces insulin sensitivity.

The findings that systemic insulin M1resistance ameliorated bv macrophage depletion and aggravated by adoptive transfer of M2 to M1 cells strongly suggest that CD206(+) cells are regarded as anti-inflammatory cells and has an impact on systemic metabolism [23-26]. We, however, obtained reverse results showing that depletion of CD206(+) cells resulted in improved glucose tolerance rather than worsening glucose metabolism. We CD206 demonstrated (M2-like) that macrophage may be responsible to regulate insulin resistance and metabolic disorders indicating that M2-like macrophages itself does not furnish insulin sensitivity but just a reflection of favorable environment. This finding led us to look for other homeostatic functions of ATMs than previously reported their anti-inflammatory function. We found that partial depletion of

CD206(+) cells lead to increase the number of smaller adipocytes with expression of metabolic genes characteristics of better metabolic function such as PGC-1α. In addition, we also found an increased number of PDGFRα+ adipocyte progenitors that are differentiated into adipocytes. These data suggested that CD206(+) macrophages in adipose tissue inhibit uncontrolled proliferation of adipocyte progenitors and excessive differentiation into mature adipocytes.

So far how adipocyte progenitor's proliferation and their differentiation into mature adipocytes in vivo is regulated is largely unknown. Our data demonstrated that adipose tissue M2-like macrophage is able to keep adipocyte progenitors in a hibernating state to inhibit excessive proliferation. This is somewhat similar to the previous report that nonmyelinating Schwann cells (HSCs) inhibit the growth of hematopoietic stem cells in the bone marrow by activating TGFB pathways [27] in the bone marrow by activating TGFβ pathways. Like an HSCs riche in the bone marrow, it may be possible that CD206(+) cells form a niche for adipocyte progenitors, thus maintaining the number of adipocyte progenitors and matured adipocyte by inhibiting the proliferation of progenitors. When the function of CD206(+) cells is abruptly compromised, the progenitor activation and differentiation into adipocytes occur. Recently, Lee and Granneman reported that β3-agonist treatment early several days resulted in adipocyte cell death, engulfment of all debris by macrophages which express the canonical markers of alternatively activated macrophages (AAMs), and recruitment of PDGFRα(+) adipocyte progenitors [28, 29].

Previous reports have shown that signals or stimuli that activate alternative activation such as extrinsic eosinophil-IL-4/-13 pathway and intrinsic PPARγ, stat6 pathway improve adipocyte function, thus leading to insulin sensitivity. Most of the experiments were performed under HFD conditions and

demonstrated that AAMs play a role in metabolic adaptations to excessive nutrient intake through anti-inflammatory functions. How should our results be reconciled with preciously reported data? First, most of the experiments were conducted on HFD condition, and adipose tissue resident macrophages may be required for metabolic adaptations to excessive nutrient intake. Second, almost all the previous reports utilized mice with genetic disruption of genes involved in alternative activation pathway. Those mice have some adaptation during both development and postnatal stage.

Our study demonstrated that CD206(+) cells inhibit progenitor proliferation and differentiation into adipocytes in a TGFBdependent manner. Further research evidences are required to expand and identify the role of TGFB in controlling the progenitor proliferation. It is recently reported that systemic lack of TGFB signal resulted in an insulin-sensitive and metabolically healthy with phenotype increased browning phenomenon in inguinal WAT. However, we observed that browning of WAT was enhanced after depleting CD206(+) cells.

CONCLUSION

The results described here unfold a fresh perspective to relate the role of M2-like macrophages in insulin resistance and metabolic disorders. We identify the novel role of ATMs to inhibit progenitor proliferation and maturation into adipocytes, thus marinating adiposity and insulin sensitivity of the whole body. This observation will shed a light on not only the mechanism of adipocyte progenitor physiology but also the role of adipose tissue-resident macrophages to regulate systemic glucose metabolism.

REFERENCES

- Sung HK, Doh KO, Son JE, Park JG, Bae Y, Choi S, et al. Adipose vascular endothelial growth factor regulates metabolic homeostasis through angiogenesis. Cell Metab. 2013; 17(1):61-72.
- 2. Sun K, Kusminski CM, Scherer PE. Adipose tissue remodeling and obesity. J Clin Invest. 2011; 121(6):2094-101.
- Hosogai N, Fukuhara A, Oshima K, Miyata Y, Tanaka S, Segawa K, Furukawa S, et al. Adipose tissue hypoxia in obesity and its impact on adipocytokine dysregulation. Diabetes. 2007; 56(4):901-11.
- Halberg N, Khan T, Trujillo ME, Wernstedt-Asterholm I, Attie AD, Sherwani S, et al. Hypoxia-inducible factor 1α induces fibrosis and insulin resistance in white adipose tissue. Mol Cell Biol. 2009; 29(16):4467-83.
- 5. Ke Q,Costa M. Hypoxia-inducible factor-1 (HIF-1). Mol Pharmacol. 2006;70(5):1469-80.
- Park YS, David AE, Huang Y, Park JB, He H, Byun Y, et al. In vivo delivery of cell-permeable antisense hypoxia-inducible factor 1α oligonucleotide to adipose tissue reduces adiposity in obese mice. J Control Release 2012; 161(1):1-9.
- Sun K, Halberg N, Khan M, Magalang UJ, Scherer PE. Selective inhibition of hypoxiainducible factor 1alpha ameliorates adipose tissue dysfunction. Mol Cell Biol 2013; 33:904-17.
- Shin MK, Drager LF, Yao Q, Bevans-Fonti S, Yoo DY, Jun JC, et al. Metabolic consequences of high-fat diet are attenuated by suppression of HIF-1α. PLoS One. 2012; 7(10):e46562.
- Zhang X, Lam KS, Ye H, Chung SK, Zhou M, Wang Y, et al. Adipose tissue-specific inhibition of hypoxia-inducible factor 1{alpha} induces obesity and glucose intolerance by impeding energy expenditure in mice. J Biol Chem. 2010;285(43):32869-77.
- Jiang C, Qu A, Matsubara T, Chanturiya T, Jou W, Gavrilova O, et al. Disruption of hypoxiainducible factor 1 in adipocytes improves insulin

- sensitivity and decreases adiposity in high-fat diet-fed mice. Diabetes. 2011; 60(10):2484-95.
- Cramer T, Yamanishi Y, Clausen BE, Förster I, Pawlinski R, Mackman N, et al. HIF-1alpha is essential for myeloid cell-mediated inflammation. Cell. 2003; 112(5):645-57.
- Fujisaka S, Usui I, Ikutani M, Aminuddin A, Takikawa A, Tsuneyama K, et al. Adipose tissue hypoxia induces inflammatory M1 polarity of macrophages in an HIF-1α-dependent and HIF-1α-independent manner in obese mice. Diabetologia. 2013; 56(6):1403-12.
- Lewis C, Murdoch C. Macrophage responses to hypoxia: implications for tumor progression and anti-cancer therapies. Am J Pathol. 2005; 167(3):627-35.
- 14. Sun K, Asterholm IW, Kusminski CM, Bueno AC, Wang ZV, Pollard JW, et al. Dichotomous effects of VEGF-A on adipose tissue dysfunction. Proc Natl Acad Sci. 2012; 109(15):5874-9.
- 15. Li J, Yu X, Pan W, Unger RH. Gene expression profile of rat adipose tissue at the onset of high-fat-diet obesity. Am J Physiol Endocrinol Metab. 2002; 282(6):E1334-41.
- Castellot JJ, Karnovsky MJ, Spiegelman BM. Differentiation-dependent stimulation of neovascularization and endothelial cell chemotaxis by 3T3 adipocytes. Proc Natl Acad Sci. 1982; 79(18):5597-601.
- 17. Takikawa A, Mahmood A, Nawaz A, Kado T, Okabe K, Yamamoto S, *et al.* HIF-1α in myeloid cells promotes adipose tissue remodeling toward insulin resistance. Diabetes. 2016; db160012.
- Olefsky JM, Glass CK. Macrophages, inflammation, and insulin resistance. Annu Rev Physiol. 2010; 72:219-46.
- Fujisaka S, Usui I, Bukhari A, Ikutani M, Oya T, Kanatani Y, et al. Regulatory mechanisms for adipose tissue M1 and M2 macrophages in dietinduced obese mice. Diabetes. 2009; 58(11): 2574-82.
- Saito M, Iwawaki T, Taya C, Yonekawa H, Noda M, Inui Y, et al. Diphtheria toxin receptor mediated conditional and targeted cell ablation

- in transgenic mice. Nat Biotechnol. 2001;19(8):746.
- 21. Patsouris D, Li PP, Thapar D, Chapman J, Olefsky JM, Neels JG. Ablation of CD11c-positive cells normalizes insulin sensitivity in obese insulin resistant animals. Cell Metab. 2008; 8(4):301-9.
- 22. Nawaz A, Aminuddin A, Kado T, Takikawa A, Yamamoto S, Tsuneyama K, et al. CD206+ M2-like macrophages regulate systemic glucose metabolism by inhibiting proliferation of adipocyte progenitors. Nat Commun. 2017; 8(1):286.
- 23. Qatanani M, Lazar MA. Mechanisms of obesity-associated insulin resistance: many choices on the menu. Genes Dev. 2007; 21(12):1443-55.
- 24. Odegaard JI, Ricardo-Gonzalez RR, Eagle AR, Vats D, Morel CR, Goforth MH, et al. Alternative M2 activation of Kupffer cells by PPARδ ameliorates obesity-induced insulin resistance. Cell Metab. 2008 Jun 4;7(6):496-507.
- 25. Pollard JW. Trophic macrophages in development and disease. Nat Rev Immunol. 2009; 9(4):259.

- 26. Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. Nature. 2013; 496(7446):445.
- 27. Yamazaki S, Ema H, Karlsson G, Yamaguchi T, Miyoshi H, Shioda S, et al. Nonmyelinating Schwann cells maintain hematopoietic stem cell hibernation in the bone marrow niche. Cell. 2011; 147(5):1146-58.
- 28. Lee YH, Kim SN, Kwon HJ, Maddipati KR, Granneman JG. Adipogenic role of alternatively activated macrophages in β-adrenergic remodeling of white adipose tissue. American Journal of Physiology-Regulatory, Integrative and Comparative Physiology. 2015; 310(1):R55-65.
- 29. Lee YH, Thacker R, Hall B, Kong R, Granneman JG. Exploring the activated adipogenic niche: Interactions of macrophages and adipocyte progenitors. Cell Cycle. 2014; 13(2):184-90.